Appendix A. MATCHMAKER One-Hybrid Vectors

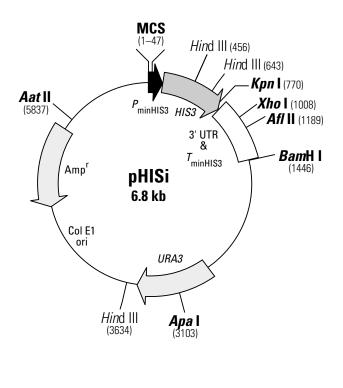


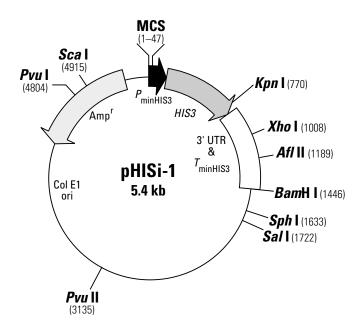


Figure 5. Map and multiple cloning site (MCS) of pHISi. pHISi is a yeast integration and reporter vector for use with the MATCHMAKER One-Hybrid System. pHISi contains the yeast HIS3 gene downstream of the MCS and the minimal promoter of the HIS3 locus (P_{minHIS}). Cis-acting sequences of interest (i.e., target elements) can be inserted into the MCS. Without activation by a target element, constitutive HIS3 expression from P_{minHIS} is very low in yeast, but allows enough growth to select for integration when constructing HIS3 reporter strains. During library screening, the leaky expression of HIS3 is controlled by adding 3-AT to the medium.

The yeast *URA3* and *HIS3* genes of pHISi can be used as selectable markers for integration into the nonfunctional *ura3* and *his3* loci, respectively, of the YM4271 host strain. Before integrating, the vector is linearized at the *Xho* I or *Afl* II sites (*his3* locus) or at the *Apa* I site (*ura3* locus). The *Kpn* I site cannot be used for integration because it cuts within the coding region of the *HIS3* gene, and that region is deleted in YM4271. pHISi cannot replicate autonomously in yeast. The plasmid contains a bacterial Col E1 origin (ori) and the ampicillin resistance gene (Ampr) for propagation and selection in *E. coli*. Unique restriction sites are in bold.

Technical Service

Appendix A. MATCHMAKER One-Hybrid Vectors continued



GAATTCCCGGGGGAGCTCACGCGTTCGCGAATCGATCCGCGGTCTAGA

EcoR | Smal/ Sac | Mlu | Sac | Xba |

Xma |

Figure 6. Map and multiple cloning site (MCS) of pHISi-1. pHISi-1 is a yeast integration and reporter vector for use with the MATCHMAKER One-Hybrid System. pHISi-1 contains the yeast HIS3 gene downstream of the MCS and the minimal promoter of the HIS3 locus (P_{minHIS}). Cis-acting sequences of interest (i.e., target elements) can be inserted into the MCS. Without activation by a target element, constitutive HIS3 expression from P_{minHIS} is very low in yeast, but allows enough growth to select for integration when constructing HIS3 reporter strains. During library screening, the leaky expression of HIS3 is controlled by adding 3-AT to the medium. pHISi-1 was constructed by transferring the HIS reporter gene from pHISi to the EcoRI/BamHI sites of pBR322. Leaky HIS3 expression in pHISi-1 is generally lower than that in pHISi, presumably due to differences in the flanking vector sequence.

The yeast *HIS3* gene is used as a selectable marker for integration into the nonfunctional *his3* locus of the YM4271 host strain after linearizing the vector at the *Xho* I or *AfI*II sites. The *Kpn*I site cannot be used for integration because it cuts within the coding region of the *HIS3* gene, and that region is deleted in YM4271. Because it does not carry the *URA3* marker, pHISi-1 can be used (together with pLacZi) to construct a dual *HIS3/lacZ* reporter strain. pHISi-1 cannot replicate autonomously in yeast. pHISi-1 contains a bacterial Col E1 origin (ori) and the ampicillin resistance gene (Amp^r) for propagation and selection in *E. coli*. Unique restriction sites are in bold.

Appendix A. MATCHMAKER One-Hybrid Vectors continued

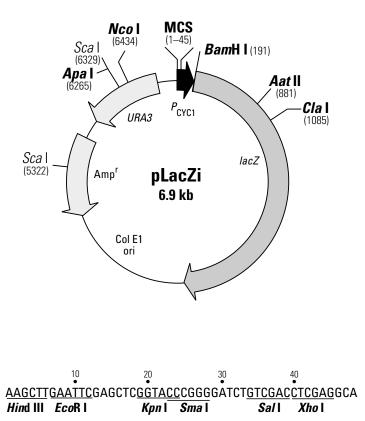


Figure 7. Map and multiple cloning site (MCS) of pLacZi. pLacZi is a yeast integration and reporter vector for use with the MATCHMAKER One-Hybrid System. This plasmid contains the bacterial lacZ gene downstream of the minimal promoter of the yeast iso-1-cytochrome C gene (P_{CYC1}). Target elements can be inserted into the MCS upstream of the P_{CYC1} -lacZ reporter. Without activation from a cis-regulatory element, lacZ expression is very low when the vector is integrated into the yeast genome. The yeast URA3 gene is used as a selectable marker for integration into the nonfunctional ura locus of the YM4271 host strain after linearizing the vector at the Nco I or Apa I site. pLacZi cannot replicate autonomously in yeast. This plasmid contains the ampicillin resistance gene (Ampr) and the Col E1 origin for selection and propagation in E. coli. Unique restriction sites are in bold.